

University of Dundee

Elevated levels of IL-6 and CRP predict the need for mechanical ventilation in COVID-19

Herold, Tobias; Jurinovic, Vindi; Arnreich, Chiara; Lipworth, Brian J.; Hellmuth, Johannes C.; von Bergwelt-Baildon, Michael

Published in:
Journal of Allergy and Clinical Immunology

DOI:
[10.1016/j.jaci.2020.05.008](https://doi.org/10.1016/j.jaci.2020.05.008)

Publication date:
2020

Document Version
Peer reviewed version

[Link to publication in Discovery Research Portal](#)

Citation for published version (APA):

Herold, T., Jurinovic, V., Arnreich, C., Lipworth, B. J., Hellmuth, J. C., von Bergwelt-Baildon, M., Klein, M., & Weinberger, T. (2020). Elevated levels of IL-6 and CRP predict the need for mechanical ventilation in COVID-19. *Journal of Allergy and Clinical Immunology*, 146(1), 128-136.e4. <https://doi.org/10.1016/j.jaci.2020.05.008>

General rights

Copyright and moral rights for the publications made accessible in Discovery Research Portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from Discovery Research Portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain.
- You may freely distribute the URL identifying the publication in the public portal.

Take down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Journal Pre-proof

Elevated levels of interleukin-6 and CRP predict the need for mechanical ventilation in COVID-19

Tobias Herold, MD, Vindi Jurinovic, PhD, Chiara Arnreich, Brian J. Lipworth, MD, Johannes C. Hellmuth, MD, Michael von Bergwelt-Baildon, MD PhD, Matthias Klein, MD, Tobias Weinberger, MD

PII: S0091-6749(20)30685-0

DOI: <https://doi.org/10.1016/j.jaci.2020.05.008>

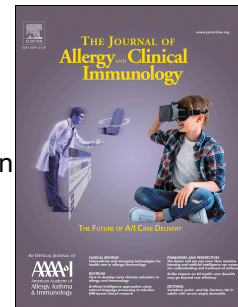
Reference: YMAI 14569

To appear in: *Journal of Allergy and Clinical Immunology*

Received Date: 28 April 2020

Revised Date: 7 May 2020

Accepted Date: 13 May 2020



Please cite this article as: Herold T, Jurinovic V, Arnreich C, Lipworth BJ, Hellmuth JC, von Bergwelt-Baildon M, Klein M, Weinberger T, Elevated levels of interleukin-6 and CRP predict the need for mechanical ventilation in COVID-19, *Journal of Allergy and Clinical Immunology* (2020), doi: <https://doi.org/10.1016/j.jaci.2020.05.008>.

This is a PDF file of an article that has undergone enhancements after acceptance, such as the addition of a cover page and metadata, and formatting for readability, but it is not yet the definitive version of record. This version will undergo additional copyediting, typesetting and review before it is published in its final form, but we are providing this version to give early visibility of the article. Please note that, during the production process, errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

© 2020 Published by Elsevier Inc. on behalf of the American Academy of Allergy, Asthma & Immunology.
This manuscript version is made available under the CC-BY-NC-ND 4.0 license
<http://creativecommons.org/licenses/by-nc-nd/4.0/>

Elevated levels of interleukin-6 and CRP predict the need for mechanical ventilation in COVID-19

Tobias Herold MD^{1,2,3,4*}, Vindi Jurinovic PhD^{1,5*}, Chiara Arnreich^{1,4}, Brian J Lipworth MD⁶, Johannes C. Hellmuth MD^{1,7}, Michael von Bergwelt-Baildon MD PhD^{1,3,8}, Matthias Klein MD^{2,9} and Tobias Weinberger MD^{2,10,11}

1 Department of Medicine III, University Hospital, LMU Munich, Munich, Germany

2 Emergency Department, University Hospital, LMU Munich, Munich, Germany

3 Nine-i Investigator

4 Research Unit Apoptosis in Hematopoietic Stem Cells, Helmholtz Zentrum München, German Center for Environmental Health (HMGU), Munich, Germany

5 Institute for Medical Information Processing, Biometry and Epidemiology, LMU Munich, Munich, Germany

6 Scottish Centre for Respiratory Research, Ninewells Hospital and Medical School, University of Dundee, Scotland, UK

7 COVID-19 Registry of the Ludwig-Maximilian-University Hospital Munich (CORKUM), University Hospital, LMU Munich, Munich, Germany

8 Intensive Care in Hematologic and Oncologic Patients (iCHOP), Cologne, Germany

9 Department of Neurology, University Hospital, LMU Munich, Munich, Germany

10 Department of Medicine I, University Hospital, LMU Munich, Munich, Germany

11 DZHK (German Centre for Cardiovascular Research), partner site Munich Heart Alliance, 80802 Munich, Germany

* Contributed equally

Corresponding Author:

Tobias Herold, MD; Marchioninstr. 15; 81377 Munich; Germany; Phone: +49 89 4400-44877; FAX: +49 89 4400-74242; Email: tobias.herold@med.uni-muenchen.de

Abstract Word count: 247/250

Article Word count: 3053/3500

Figures/Tables: 5 Figures, 3 Tables /8

Sources of funding: none

Author contributions

T.H. and T.W. conceived and designed the study. T.H., C.A., M. K. and T.W. were responsible for clinical care and collected patient data. J.C.H. was responsible for the ethical approval of the study. Statistical analysis was conducted by V.J.. M.v.B.-B. supervised all aspects of the study. B.L. corrected and helped write the manuscript and added important aspects to the analysis. T.H. and T.W. wrote the first draft. All authors contributed to data interpretation, critical revision of the manuscript and approved the final version of the manuscript.

Competing interests

B. L. reports grants and personal fees from Sanofi, AstraZeneca, and Teva; reports personal fees from Cipla, Glenmark, and Lupin; reports grants, personal fees, and other from Chiesi, outside the submitted work; and reports that his son is an employee of AstraZeneca.

M.v.B.-B. is the local principal investigator of the currently conducted COVACTA-Trial (A Study to Evaluate the Safety and Efficacy of Tocilizumab in Patients With Severe COVID-19 Pneumonia; NCT04320615, Sponsor: Hoffmann-La Roche). He has previously received honoraria and research funding from Hoffman-La Roche unrelated to this project.

47 M. K received speakers' fees from BioMerieux and served on the advisory board of
48 BioMerieux.

49 The other authors declare no conflict of interest.

50

Abstract

Background: COVID-19 can manifest as a viral induced hyperinflammation with multi-organ involvement. Such patients often experience rapid deterioration and need for mechanical ventilation. Currently, no prospectively validated biomarker of impending respiratory failure is available.

Objective: We aimed to identify and prospectively validate biomarkers that allow the identification of patients in need of impending mechanical ventilation.

Methods: Patients with COVID-19 hospitalized from February 29th to April 09th, 2020 were analyzed for baseline clinical and laboratory findings at admission and during the disease. Data from 89 evaluable patients were available for the purpose of analysis comprising an initial evaluation cohort (n=40) followed by a temporally separated validation cohort (n=49).

Results: We identified markers of inflammation, LDH and creatinine as most predictive variables of respiratory failure in the evaluation cohort. Maximal interleukin-6 (IL-6) levels before intubation showed the strongest association with the need of mechanical ventilation followed by maximal CRP. Respective AUC values for IL-6 and CRP in the evaluation cohort were 0.97 and 0.86 and similar in the validation cohort 0.90 and 0.83. The calculated optimal cutoff values in the course of disease from the evaluation cohort (IL-6 > 80 pg/ml and CRP > 97 mg/l) both correctly classified 80% of patients in the validation cohort regarding their risk of respiratory failure.

Conclusion: Maximal levels of IL-6 followed by CRP were highly predictive of the need for mechanical ventilation. This suggests the possibility of using IL-6 or CRP levels to guide escalation of treatment in patients with COVID-19 related hyperinflammatory syndrome.

Clinical Implications: IL-6 followed by CRP strongly predicted patients at risk of respiratory deterioration and might be pivotal for risk-adapted escalation of treatment.

Capsule summary: We studied laboratory parameters as predictors of impending respiratory failure in COVID-19. Maximum levels of interleukin-6 over the course of disease, followed by CRP, were the best predictors of respiratory failure in two separate cohorts.

Key words: Interleukin-6, IL-6, CRP, COVID-19, respiratory failure, mechanical ventilation, prediction, hyperinflammation

Abbreviations: COVID-19: Coronavirus Disease 2019; SARS-CoV-2: Severe Acute Respiratory Syndrome coronavirus 2; SARS: severe acute respiratory syndrome; H7N9: avian-origin influenza; H1N1: influenza A; BAL: Bronchoalveolar lavage; ROC: Receiver operating characteristic; AUC: Area under the curve; CI: Confidence interval; BMI: Body mass index; CT: Computed Tomography; CRP: C-Reactive Protein; WBC: White blood cell count; LDH: Lactate Dehydrogenase; PCT: Procalcitonin; IL6: Interleukin-6; qSOFA score: quick sequential organ failure assessment score - predicts mortality in sepsis; CURB-65 score: predicts mortality in community-acquired pneumonia; MuLBSTA score: predicts mortality in patients with viral pneumonia; q-values represent the Benjamini-Hochberg adjusted p-values

Introduction

The pandemic Coronavirus-disease 19 (COVID-19) is characterized by a highly variable course. While most patients experience only mild symptoms, a relevant proportion develops severe disease progression up to respiratory failure. Interestingly, many patients do not show signs of respiratory distress, despite severe hypoxemia in blood gas analysis ¹. About 5% of patients require intensive care including mechanical ventilation ^{2,3}.

Recently published large retrospective analyses provide a detailed characterization of COVID-19 and identify variables associated with disease severity and high mortality ^{4,5}. One of the largest studies so far shows that age, quick sequential organ failure assessment score (qSOFA score) and D-Dimer correlate with in-hospital death in a multivariate analysis ². Another group showed a correlation of obesity and increased inflammatory markers in the blood with respiratory failure ⁶.

In many aspects, severe COVID-19 may be regarded as a viral induced hyperinflammatory condition with multi-organ involvement due to a cytokine cascade ⁷. Of these various cytokines, the presence of raised circulating levels of interleukin-6 (IL-6) appears to be key and is closely connected to disease severity not only in COVID-19 ⁸ but also in avian-origin H7N9 influenza infections ⁹ and the common seasonal H1N1 influenza A ¹⁰.

While these studies identify the correlation of parameters with disease severity, prospective factors predicting impending deterioration of patients are not yet established. The broad spectrum of the disease courses and silent hypoxia make identification of patients at risk difficult. We aimed to identify variables that allow the prediction of COVID-19 patients with a high risk of respiratory failure.

Methods

Patients and study design

All patients with PCR proven COVID-19 hospitalized at our institution from February 29th to April 09th, 2020 (n=115) were screened and analyzed for baseline clinical and laboratory findings. In total, 26 patients were excluded from the study and the depicted cohort consisted of 89 patients (Table 1). Patients with palliative treatment (n=3) or hospitalization due to other medical reasons and nosocomial Sars-CoV2-infection on the ward (n=13) were excluded from this study. Additionally, patients already mechanically ventilated at admission (n=8) and those receiving anti-IL-6 antibody treatment (n=2) were excluded (Figure 1).

Of the 89 evaluable patients, 40 were part of an initial evaluation cohort hospitalized from February 29th to March 27th, 2020 (Supplementary Table E1). This cohort was used to identify predictive markers of respiratory failure.

Following an interim analysis of the initial evaluation cohort¹¹, we performed a power analysis to estimate the number of patients needed to validate our findings. Assuming the need of mechanical ventilation to be 20% in the validation cohort and the risks for mechanical ventilation to be 70% and 20% in the high-risk and the low-risk group, respectively, the total sample size for a two-sided test was determined to be 40. We defined an additional safety margin of 10%. This subsequent validation cohort consisted of patients hospitalized from March 27th to April 09th, 2020 (n=49) (Supplementary Table E2). Follow up for all patients was complete through April 12th, 2020. A comparison of both cohorts is shown in Supplementary Table E3.

Use of compassionate medication was low in the study cohort before mechanical ventilation (5 patients received lopinavir/ritonavir, 8 patients received hydroxychloroquine).

Decision on endotracheal intubation was made following internationally accepted recommendations (PaO₂/FiO₂ <150mmHg or <200mmHg in case of anticipated difficult airway)¹².

Patients are part of the COVID-19 Registry of the Ludwig-Maximilian-University Hospital Munich (CORKUM). Patient data were anonymized for analysis and the study was approved by the local ethics committee (Ethics committee of the LMU Munich, No: 20-245).

IL-6 and CRP measures

The fully automated Elecsys® system on a cobas e801 platform (Roche Diagnostics, Switzerland) was used to measure single levels of IL-6, as described previously^{13, 14}. The Elecsys® IL-6 immunoassay has been standardized against the NIBSC 1st IS 89/548 Standard. CRP values were measured on a cobas c702 platform using the Tina-quant® C-Reactive Protein assay (Roche Diagnostics, Switzerland).

Statistical analysis

All variables with less than 50% of missing data in the initial cohort were tested for the association with respiratory failure. Categorical variables were tested with the χ^2 test, and numerical variables with the Mann-Whitney U test. When appropriate, a paired test was performed. All tests were two-sided. The p-values were adjusted for multiple testing with the Benjamini-Hochberg-method to avoid inflating the alpha error. An adjusted p-value (q-value) of ≤ 0.05 was considered significant. We constructed receiver operating characteristic (ROC) curves and calculated the area under the curve (AUC) to compare the predictive ability of continuous variables. The AUC can be interpreted as the probability that the predictor's value for a randomly chosen patient requiring intubation will be higher than its value for a randomly chosen patient not requiring intubation. The optimal cut off was defined as the one maximizing the Youden's Index¹⁵. Statistical analyses were performed using the R software package (version 3.6.2). Figures were drawn using Graphpad Prism® (Version 6.0).

Results

Initial identification of IL-6 and CRP as strongest predictors of respiratory failure

To initially evaluate predictors of respiratory failure, 40 patients with confirmed COVID-19 were recruited from February 29th to March 27th, 2020 and served as an evaluation cohort (Figure 1). Thirteen (32.5%) patients deteriorated during hospitalization and required mechanical ventilation. The time from hospital admission to intubation varied from less than two hours to 9 days (median 2 days). Patients requiring mechanical ventilation did not differ in age, comorbidities, radiological findings, respiratory rate or qSofa score (Supplementary Table E1).

Heart rate, markers of inflammation, LDH and creatinine at admission were significantly associated with respiratory failure (Supplementary Table E1). Elevated IL-6 showed the strongest association with the need for mechanical ventilation (Figure 2A, $p=1.2 \times 10^{-5}$).

In addition to values at first assessment, follow-up data were available for laboratory variables. These follow-up data were used to test if there are critical laboratory values that are associated with respiratory failure once they have been reached during disease course. For each patient, we assessed the maximum level of each parameter during disease (for patients requiring ventilation, only values before intubation were used). The maximal values were correlated with respiratory failure (Table 2). Maximal IL-6 levels predicted respiratory failure with highest accuracy (Figure 2, AUC=0.97, CI [0.93, 1.0]), followed by CRP (Figure 3 AUC=0.86, CI [0.74, 0.98]) and creatinine (AUC=0.85, CI [0.74, 0.97]). The optimal cutoff for maximal IL-6 was 80 pg/ml. After reaching an IL-6 value of 80 pg/ml, the median time to mechanical ventilation was 1.5 days (range 0–4 days). The optimal cutoff for maximal CRP was 97 mg/l, with the median time to mechanical ventilation of 0 days after reaching the cutoff (range 0–4 days).

Prospective validation of calculated cutoffs for IL-6 and CRP

A cohort of 40 patients was estimated to have an adequate power to validate our findings (see Methods). The validation cohort prospectively recruited 49 patients from March 27th to April 09th, 2020, of which 19 (39%) required mechanical ventilation. As in the initial cohort, creatinine, LDH, and several markers of inflammation were significantly elevated in patients requiring intubation (Table 2 and Supplementary Table E2). Again, IL-6 at assessment was strongly associated with respiratory failure (Figure 2B), and maximal IL-6 was the best predictor of future respiratory failure among all parameters (Figure 2D, AUC 0.90, CI [0.81, 0.98], Table 2). CRP values at initial assessment were significantly associated with respiratory failure (Figure 2F and Figure 3 AUC=0.86, CI [0.75, 0.96]). Follow-up values of CRP during the disease course did not improve the prediction of respiratory failure in the validation cohort (Table 2, AUC=0.83, CI [0.72, 0.95]).

To validate our findings from the initial cohort, we analyzed the number of patients correctly classified regarding their need of mechanical respiratory support by the determined cutoffs of IL-6 and CRP at presentation and in the course of disease (Table 3). At presentation, IL-6 >35 pg/ml as well as CRP >32.5 mg/l showed high sensitivity to detect patients at risk for respiratory failure (84% and 95%) with moderate specificity (63% for both parameters). Measuring IL-6 and CRP values in the course of disease (cutoffs 80 pg/ml and 97 mg/l) increased the specificity for both parameters (83% and 77%) accompanied with a decrease in sensitivity (74% vs. 84%). In detail, nineteen (39%) patients exceeded the calculated maximal IL-6 cutoff (>80 pg/ml) in the validation cohort, compared to 23 (47%) patients exceeding the CRP cutoff (>97mg/l). Of these patients, 74% and 70% were correctly classified by IL-6 and CRP, respectively, as being at risk for respiratory failure (positive predictive value). Of the 30 patients with values below the IL-6 cutoff, 83% did not require mechanical ventilation, while this was the case for 88% of the 26 patients remaining below the CRP cutoff of 97 mg/l (negative predictive value). In total, the calculated cutoffs for maximal IL-6 and CRP both correctly classified 80% of patients regarding their risk of respiratory failure (Table 3), while values at assessment show poorer predictor properties owing to the moderate specificity (correct classification of 71% for IL-6 and 76% for CRP)

Taken together, while both values have a strong sensitivity at assessment, specificity is gained when examining values in the course of disease. The risk ratios for the cutoffs of IL-6 and CRP were 4.4 and 6.0 in the validation cohort, with corresponding p-values of 0.00022 and 0.00011. The optimal cut point in the validation cohort was slightly lower for IL-6 (60 pg/ml) and identical for CRP (97 mg/l).

Predictive values of the combined cohort

To further evaluate positive and negative predictive values (PPV/NPV) of IL-6 and CRP we combined the two cohorts (Table 1). We calculated predictive values across the range of all possible cutoffs. The PPV of CRP was consistently lower compared to IL-6 in the overall study cohort (Figure 4). In other words, increased CRP misclassified more patients as being at risk for respiratory failure than IL-6. However, the predictive values strongly depend on the selected cutoff (Figure 4). For cutoffs <50 pg/ml for IL-6 and <40 mg/l for CRP (dotted line), the risk of intubation for patients with sub-threshold levels is roughly zero, while patients with levels above these values show a dramatic increase in the risk of respiratory failure. The risk for respiratory failure in patients with IL-6 levels exceeding 210 pg/ml was 100% (dashed line). The NPV of IL-6 and CRP parameters was comparable. In the combined cohort, the optimal threshold value (maximal Youden index¹⁵) is highest at 65 pg/ml for IL-6 and for CRP at 97 mg/l (corresponding risk ratio of 18.1 and 6.9).

Furthermore, we analyzed the time lag from reaching the cutoff values to intubation in the combined cohort. Patients reached the cutoff of IL-6 (>65 ng/ml) and CRP (>97 mg/l) at a median of 23.2 and 15.7 hours before intubation, resulting in a significant time difference between the two values of 7.5 hours in favor for IL-6 (Figure 5; p=0.014).

Discussion

Our study in hospitalized patients with COVID-19 has provided three key findings: First, circulating levels of IL-6 as well as CRP were highly predictive of the need for invasive ventilation, with corresponding AUC values of 0.97 and 0.90 for IL-6 and 0.86 and 0.83 for CRP in the first and the second cohorts, respectively. Secondly, we defined cutoffs for IL-6 (at presentation >35 pg/ml; maximal value >80 pg/ml) and CRP (at presentation >32.5 mg/l; maximal value >97 mg/l) in the evaluation cohort. Cutoff values at assessment correctly classified 71% (for IL-6) and 76% (for CRP) of patients in the validation cohort with a further increase when measuring maximal values in the course of disease (80% for both parameters). Thirdly, elevated IL-6 levels in the course of disease predicted respiratory failure significantly earlier than CRP (23.2 vs. 15.7 hours). Therefore, IL-6 and CRP are useful markers that predict impending respiratory failure with high accuracy and can help physicians correctly allocate patients who might benefit from early treatment escalation, for example using anti-cytokine strategies. We believe that having these data reproduced across the two separate cohorts enhances the strength of our conclusions. It is important to note that the commercial diagnostic IL-6 assay used in our study allows the measurement of IL-6 in a comparable time scale as CRP. Since it uses the broadly available Cobas platform it can be implemented in most laboratories.

Our study also has several limitations. It is still unclear whether elevated inflammatory markers merely represent an epiphenomenon or a causal pathogenic element of severe COVID-19¹⁶. It is likely that elevated IL-6 reflects the cytokine mediated hyperinflammatory state as evidenced by the similarly predictive values for CRP. Further, even though IL-6 and CRP levels are significantly elevated in patients requiring ventilation, they are relatively low compared to levels observed in patients with septic shock¹⁷. However, earlier studies in severe acute respiratory syndrome (SARS) or H7N9 influenza patients show that inflammatory cytokines are highly expressed in lung tissues. Autopsy reports from SARS patients showed a high amount of inflammatory cytokines in cells expressing angiotensin-

converting enzyme 2¹⁸, the functional receptor for SARS-CoV and in even higher affinity for SARS-CoV2¹⁹. Bronchoalveolar lavage (BAL) in H7N9 influenza patients showed 10³ times higher concentrations of different cytokines including IL-6 compared to plasma levels, hinting towards a massively increased local concentration of inflammatory cytokines in the diseased lung⁹. Recent preprints provide detailed single cell RNA-sequencing data from immune cells in peripheral blood as well as BAL from COVID-19 patients. The authors report that peripheral monocytes did not substantially express proinflammatory cytokines²⁰, while there was high expression in monocyte derived macrophages in BAL²¹. Taken together, these data possibly suggest that circulating levels of IL-6 might be a putative surrogate for the burden of lung tissue damage and provide a “window” into the lung⁹.

IL-6 and CRP have been associated with severity of COVID-19 (in most cases defined by the Chinese National Health Commission) and mortality before²²⁻²⁴. However, to our knowledge our study is the first to demonstrate a prospective prediction of the end point “mechanical ventilation”, which is of high clinical relevance not only for patient treatment but also for resource planning. Very recent publications provide additional data that strengthen the role of IL-6 and CRP in COVID-19 as predictive markers^{22, 23}. Unfortunately, these studies did not include a prospective validation cohort and sometimes did not mention analysis platforms²². A further difference between our and other studies is the dramatic discrepancy in mortality of severely diseased patients. We are not able to analyze mortality as an end point because only two patients had died until April 12th. This number has only increased by one until May 6th (overall mortality 3.4%). While still some patients are in critical condition and the mortality rate in our cohort is likely to increase in the next weeks it will be significantly below those reported. We can only speculate about the reasons for this huge difference but argue that overwhelmed hospitals and patient selection might have contributed to the increased mortality observed in other studies. As we did not perform sequential CT-scans after 24-48 hours in our patients due to radiation hygiene, we are not able to precisely calculate severity of COVID-19 according the Chinese National Health Commission classification to compare our patient cohort to the cohorts of the mentioned studies. However, our validation cohort at

least exists of 63% of severe patients due to the available parameters (2% with mild and 35% with moderate symptoms), which exceeds the recently published cohorts^{22, 23}.

Since the start of the pandemic, hundreds of research articles on COVID-19 have been published²⁵. To our knowledge, we report the first predictive marker for respiratory failure that was prospectively validated in an independent cohort. Although our sample sizes were small, the large difference in risk for respiratory failure between the high-risk and the low-risk group made it possible to successfully validate our findings. Interestingly, a study of 134 patients with avian-origin H7N9 influenza in 2013 also showed a strong correlation of IL-6 and disease severity. In analogy to our findings, this study reports that IL-6 plasma levels >80 pg/ml were found in all patients with lethal outcome compared to only 8.3% in surviving patients⁹. The combined cohort (n=89) produced an only slightly lower cutoff for IL-6 (65 pg/ml) while the cutoff for CRP levels remained the same at 97 mg/l when calculated from the combined cohort. However, even the combined sample size is probably too small to determine an optimal cutoff value. Furthermore, the acceptable proportion of falsely identified low-risk patients, and therefore the set threshold, is largely dictated by the availability of health care resources. Future prospective studies with larger sample sizes are needed to formally address this issue. We want to stress that IL-6 and CRP should be used as a predictor not an indication for invasive respiratory support, as mechanical ventilation per se has several unintended adverse consequences and may support inflammation of distal airways in COVID-19 patients.

Immunologically, CRP and IL-6 are closely intertwined. IL-6 is known to induce gene expression and release of CRP from the liver^{26, 27} and also from immune cells²⁸. A functional connection has been shown in different trials using IL-6 inhibition, in which CRP-levels rapidly normalized after blocking IL-6²⁹. In analogy, we found that IL-6 levels predicted respiratory failure significantly earlier than CRP-levels, which is essential for a predictive marker. While inhibition of inflammatory pathways represents a promising approach to treat hyperinflammatory COVID-19 patients, inhibition of IL-6 could be detrimental in the immune

response to virus-induced pneumonias^{30, 31}. Thus, our study does not facilitate any recommendations for or against IL-6 inhibition. Ongoing randomized controlled clinical trials of IL-6-antibodies in the treatment of COVID-19 will shed light on this question (e.g. NCT04320615 and NCT04331795). More importantly, in times of missing established therapeutic options, best supportive care is essential³².

In summary, we were able to validate our finding that IL-6 and CRP levels serve as strong predictors of patients in need of ventilator support. In the current situation with overwhelmed intensive care units and overcrowded emergency rooms, correct identification of patients in need of intensive care is crucial. Assessing these parameters to identify patients at risk of respiratory failure at an early stage might be helpful for triage planning and timely allocation of critically ill patients as well as a guide to escalation of treatment strategies in COVID-19 patients.

Acknowledgements

We would like to thank all CORKUM investigators and staff. The authors thank the patients and their families for their participation in the CORKUM registry as well as all health care workers for their outstanding service.

References

1. Xie J, Tong Z, Guan X, Du B, Qiu H, Slutsky AS. Critical care crisis and some recommendations during the COVID-19 epidemic in China. *Intensive Care Med* 2020.
2. Zhou F, Yu T, Du R, Fan G, Liu Y, Liu Z, et al. Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: a retrospective cohort study. *Lancet* 2020; 395:1054-62.
3. Wang D, Hu B, Hu C, Zhu F, Liu X, Zhang J, et al. Clinical Characteristics of 138 Hospitalized Patients With 2019 Novel Coronavirus-Infected Pneumonia in Wuhan, China. *JAMA* 2020.
4. Grasselli G, Zangrillo A, Zanella A, Antonelli M, Cabrini L, Castelli A, et al. Baseline Characteristics and Outcomes of 1591 Patients Infected With SARS-CoV-2 Admitted to ICUs of the Lombardy Region, Italy. *JAMA* 2020.
5. Petrilli CM, Jones SA, Yang J, Rajagopalan H, O'Donnell LF, Chernyak Y, et al. Factors associated with hospitalization and critical illness among 4,103 patients with COVID-19 disease in New York City. *medRxiv* 2020.
6. Dreher M, Kersten A, Bickenbach J, Balfanz P, Hartmann B, Cornelissen C, et al. Charakteristik von 50 hospitalisierten COVID-19-Patienten mit und ohne ARDS. *Dtsch Arztebl International* 2020; 117:271-8.
7. Lipworth B, Chan R, Lipworth S, RuiWen Kuo C. Weathering the Cytokine Storm in Susceptible Patients with Severe SARS-CoV-2 Infection. *J Allergy Clin Immunol Pract* 2020.
8. Chen X, Zhao B, Qu Y, Chen Y, Xiong J, Feng Y, et al. Detectable serum SARS-CoV-2 viral load (RNAemia) is closely correlated with drastically elevated interleukin 6 (IL-6) level in critically ill COVID-19 patients. *Clin Infect Dis* 2020.
9. Wang Z, Zhang A, Wan Y, Liu X, Qiu C, Xi X, et al. Early hypercytokinemia is associated with interferon-induced transmembrane protein-3 dysfunction and predictive of fatal H7N9 infection. *Proc Natl Acad Sci U S A* 2014; 111:769-74.

10. Hagau N, Slavcovici A, Gongnanu DN, Oltean S, Dirzu DS, Brezoszki ES, et al. Clinical aspects and cytokine response in severe H1N1 influenza A virus infection. Crit Care 2010; 14:R203.
11. Herold T, Jurinovic V, Arnreich C, Hellmuth JC, von Bergwelt-Baildon M, Klein M, et al. Level of IL-6 predicts respiratory failure in hospitalized symptomatic COVID-19 patients. medRxiv 2020.
12. Bellani G, Laffey JG, Pham T, Fan E, Brochard L, Esteban A, et al. Epidemiology, Patterns of Care, and Mortality for Patients With Acute Respiratory Distress Syndrome in Intensive Care Units in 50 Countries. JAMA 2016; 315:788-800.
13. Fischer SK, Williams K, Wang L, Capio E, Briman M. Development of an IL-6 point-of-care assay: utility for real-time monitoring and management of cytokine release syndrome and sepsis. Bioanalysis 2019; 11:1777-85.
14. Jekarl DW, Lee SY, Lee J, Park YJ, Kim Y, Park JH, et al. Procalcitonin as a diagnostic marker and IL-6 as a prognostic marker for sepsis. Diagn Microbiol Infect Dis 2013; 75:342-7.
15. Youden WJ. Index for rating diagnostic tests. Cancer 1950; 3:32-5.
16. Hirano T, Murakami M. COVID-19: a new virus, but an old cytokine release syndrome. Immunity 2020; pre-proof print.
17. Surbatovic M, Popovic N, Vojvodic D, Milosevic I, Acimovic G, Stojicic M, et al. Cytokine profile in severe Gram-positive and Gram-negative abdominal sepsis. Sci Rep 2015; 5:11355.
18. He L, Ding Y, Zhang Q, Che X, He Y, Shen H, et al. Expression of elevated levels of pro-inflammatory cytokines in SARS-CoV-infected ACE2+ cells in SARS patients: relation to the acute lung injury and pathogenesis of SARS. J Pathol 2006; 210:288-97.
19. Shang J, Ye G, Shi K, Wan Y, Luo C, Aihara H, et al. Structural basis of receptor recognition by SARS-CoV-2. Nature 2020.

- 401 20. Wilk AJ, Rustagi A, Zhao NQ, Roque J, Martinez-Colon GJ, McKechnie JL, et al. A
402 single-cell atlas of the peripheral immune response to severe COVID-19. medRxiv
403 2020.
- 404 21. Liao M, Liu Y, Yuan J, Wen Y, Xu G, Zhao J, et al. The landscape of lung
405 bronchoalveolar immune cells in COVID-19 revealed by single-cell RNA sequencing.
406 medRxiv 2020.
- 407 22. Liu F, Li L, Xu M, Wu J, Luo D, Zhu Y, et al. Prognostic value of interleukin-6, C-
408 reactive protein, and procalcitonin in patients with COVID-19. J Clin Virol 2020;
409 127:104370.
- 410 23. Zhu Z, Cai T, Fan L, Lou K, Hua X, Huang Z, et al. Clinical value of immune-
411 inflammatory parameters to assess the severity of coronavirus disease 2019. Int J
412 Infect Dis 2020.
- 413 24. Velavan TP, Meyer CG. Mild versus severe COVID-19: laboratory markers. Int J
414 Infect Dis 2020.
- 415 25. London AJ, Kimmelman J. Against pandemic research exceptionalism. Science 2020.
- 416 26. Castell JV, Gomez-Lechon MJ, David M, Andus T, Geiger T, Trullenque R, et al.
417 Interleukin-6 is the major regulator of acute phase protein synthesis in adult human
418 hepatocytes. FEBS Lett 1989; 242:237-9.
- 419 27. Castell JV, Andus T, Kunz D, Heinrich PC. Interleukin-6. The major regulator of
420 acute-phase protein synthesis in man and rat. Ann N Y Acad Sci 1989; 557:87-99;
421 discussion 100-1.
- 422 28. Sehgal PB. Interleukin-6: a regulator of plasma protein gene expression in hepatic
423 and non-hepatic tissues. Mol Biol Med 1990; 7:117-30.
- 424 29. Nishimoto N, Terao K, Mima T, Nakahara H, Takagi N, Kakehi T. Mechanisms and
425 pathologic significances in increase in serum interleukin-6 (IL-6) and soluble IL-6
426 receptor after administration of an anti-IL-6 receptor antibody, tocilizumab, in patients
427 with rheumatoid arthritis and Castleman disease. Blood 2008; 112:3959-64.

30. Yang ML, Wang CT, Yang SJ, Leu CH, Chen SH, Wu CL, et al. IL-6 ameliorates acute lung injury in influenza virus infection. *Sci Rep* 2017; 7:43829.
31. Lauder SN, Jones E, Smart K, Bloom A, Williams AS, Hindley JP, et al. Interleukin-6 limits influenza-induced inflammation and protects against fatal lung pathology. *Eur J Immunol* 2013; 43:2613-25.
32. Clinical management of severe acute respiratory infection when Covid-19 is suspected.: World Health Organization; Updated March 13, 2020. Accessed April 14, 2020.] Available from [https://www.who.int/publications-detail/clinical-management-of-severe-acute-respiratory-infection-when-novel-coronavirus-\(ncov\)-infection-is-suspected](https://www.who.int/publications-detail/clinical-management-of-severe-acute-respiratory-infection-when-novel-coronavirus-(ncov)-infection-is-suspected).
33. Seymour CW, Liu VX, Iwashyna TJ, Brunkhorst FM, Rea TD, Scherag A, et al. Assessment of Clinical Criteria for Sepsis: For the Third International Consensus Definitions for Sepsis and Septic Shock (Sepsis-3). *JAMA* 2016; 315:762-74.
34. Lim WS, van der Eerden MM, Laing R, Boersma WG, Karalus N, Town GI, et al. Defining community acquired pneumonia severity on presentation to hospital: an international derivation and validation study. *Thorax* 2003; 58:377-82.
35. Guo L, Wei D, Zhang X, Wu Y, Li Q, Zhou M, et al. Clinical Features Predicting Mortality Risk in Patients With Viral Pneumonia: The MuLBSTA Score. *Front Microbiol* 2019; 10:2752.

448 **Tables**449 **Table 1: Combined Cohort**

Variable	Evaluable	Median (range) / n (%)	Mechanical ventilation		p-value	q-value
			No (n = 57)	Yes (n = 32)		
Baseline Characteristics *						
Age (years)	89	61 (18 - 84)	58 (18 - 84)	65 (45 - 81)	0.031	0.067
Respiratory rate (/min)	74	18 (11 - 40)	17 (13 - 39)	25 (11 – 40)	0.0024	0.0073
Heart rate (/min)	66	86 (54 - 130)	85 (54 - 130)	89 (64 - 112)	0.32	0.47
BMI	71	26.9 (18.1 – 45.7)	26.0 (18.1 – 36.2)	27.6 (18.3 – 45.7)	0.074	0.15
Male gender	89	62 (70)	33 (58)	29 (91)	0.0029	0.0073
Any comorbidities	87	70 (80)	43 (77)	27 (87)	0.38	0.53
Hypertension	86	45 (52)	25 (45)	20 (65)	0.14	0.25
Diabetes mellitus	86	13 (15)	7 (13)	6 (19)	0.61	0.68
Coronary heart disease	85	7 (8)	4 (7)	3 (10)	>0.99	>0.99
Chronic obstructive lung disease	86	9 (10)	7 (13)	2 (6)	0.54	0.67
Computed Tomography [#]						
Consolidation	78	46 (59)	30 (59)	16 (59)	>0.99	>0.99
Ground glass opacity	78	72 (92)	47 (92)	25 (93)	>0.99	>0.99
Bilateral infiltration	78	70 (90)	44 (86)	26 (96)	0.32	0.47
Scores [§]						
qSOFA score ³³	71	30 (42)	13 (28)	17 (68)	0.0028	0.0073
CURB-65 score ³⁴ ≥ 1	47	22 (47)	11 (41)	11 (55)	0.50	0.67
MuLBSTA score ³⁵	68	11 (0 - 15)	9 (0 - 15)	11 (5 - 15)	0.090	0.17
Laboratory parameters [#]	Evaluable	Median (range)	Mechanical ventilation		p-value	q-value
			No (n = 57)	Yes (n = 32)		
Lymphocyte count (G/l)	67	0.92 (0.20 – 2.84)	0.85 (0.31 – 2.36)	0.94 (0.20 – 2.84)	0.60	0.68
CRP (mg/l)	89	36 (0 - 369)	20 (0 - 315)	93 (16 - 369)	1.9·10⁻⁷	2.6·10⁻⁶
Bilirubin (mg/dl)	84	0.5 (0.2 – 1.9)	0.5 (0.2 – 1.2)	0.6 (0.2 – 1.9)	0.19	0.32
WBC (G/l)	89	5.86 (0.15 – 308)	5 (1.92 – 12.4)	7.26 (0.15 - 308)	0.0024	0.0073
LDH (U/l)	88	311 (153 - 1121)	278 (153 - 619)	462 (240 - 1121)	1.5·10⁻⁶	0.000010
PCT (ng/ml)	87	0 (0 - 5)	0 (0 – 0.6)	0.2 (0 - 5)	8.7·10⁻⁷	8.1·10⁻⁶
IL-6 (pg/ml)	86	34 (0 - 430)	23.2 (0 - 209)	95.4 (14.2 - 430)	2.3·10⁻⁹	6.5·10⁻⁸
Thrombocyte count (G/l)	89	194 (0.12 - 450)	194 (0.27 - 383)	202 (0.12 - 450)	0.55	0.67
Troponin T (ng/ml)	78	0 (0 – 0.178)	0 (0 – 0.143)	0 (0 – 0.178)	0.00010	0.00047
Creatinine (mg/dl)	89	0.9 (0.4 – 7)	0.9 (0.4 – 5.6)	1.1 (0.8 – 7)	5.2·10⁻⁶	0.000029
D-Dimer	76	0.7 (0 – 35.2)	0.6 (0 – 35)	0.9 (0 – 35.2)	0.0079	0.018
Ferritin (ng/ml)	79	703 (30 - 3577)	545 (30 - 2578)	1392 (237 - 3577)	0.00023	0.00092

450 * respiratory rate and heart rate and BMI (Body mass index) were measured at admission;
451 existing comorbidities were evaluated by patient history at admission; # CT-scans and
452 laboratory parameters at admission; § scores were calculated at admission. CRP = C-
453 Reactive Protein; WBC= White blood cell count; LDH = Lactate Dehydrogenase; PCT =
454 Procalcitonin; IL6 = Interleukin-6; qSOFA score = predicts mortality in sepsis, CURB-65
455 score = predicts mortality in community-acquired pneumonia, MuLBSTA score = predicts
456 mortality in patients with viral pneumonia; q-values represent the Benjamini-Hochberg
457 adjusted p-values

458

459

Table 2: p-values, AUC's and optimal cutoffs in evaluation, validation and combined cohort

Variable	Evaluation set						Validation set						Combined cohort					
	At presentation			Maximal			At presentation			Maximal			At presentation			Maximal		
	p-value	AUC [CI]	Cutoff	p-value	AUC [CI]	Cutoff	p-value	AUC [CI]	Cutoff	p-value	AUC [CI]	Cutoff	p-value	AUC [CI]	Cutoff	p-value	AUC [CI]	Cutoff
IL-6 pg/ml	0.000012	0.94 [0.86, 1.00]	35	$5.4 \cdot 10^{-8}$	0.97 [0.93, 1.00]	80	0.000076	0.84 [0.73, 0.95]	48.9	$4.9 \cdot 10^{-7}$	0.90 [0.81, 0.98]	60	$2.3 \cdot 10^{-9}$	0.89 [0.81, 0.96]	48.9	$2.6 \cdot 10^{-11}$	0.93 [0.88, 0.98]	65
CRP mg/l	0.0031	0.79 [0.65, 0.93]	32.5	0.00027	0.86 [0.74, 0.98]	97	0.000032	0.86 [0.75, 0.96]	32.5	0.000097	0.83 [0.72, 0.95]	97	$1.9 \cdot 10^{-7}$	0.83 [0.75, 0.92]	32.5	$7.0 \cdot 10^{-8}$	0.85 [0.76, 0.93]	97
PCT ng/ml	0.0043	0.74 [0.58, 0.90]	0.05	0.0084	0.74 [0.57, 0.91]	0.25	0.000073	0.81 [0.69, 0.93]	0.05	0.00015	0.80 [0.67, 0.93]	0.25	$8.7 \cdot 10^{-7}$	0.78 [0.68, 0.88]	0.05	$4.2 \cdot 10^{-6}$	0.78 [0.67, 0.88]	0.25
LDH U/l	0.00062	0.83 [0.70, 0.97]	320	0.071	0.68 [0.50, 0.86]	590	0.00032	0.81 [0.67, 0.95]	410	0.0076	0.73 [0.60, 0.89]	440	$1.4 \cdot 10^{-6}$	0.81 [0.72, 0.91]	410	0.0015	0.70 [0.59, 0.82]	380.5
WBC G/l	0.0028	0.80 [0.66, 0.93]	4920	0.010	0.75 [0.58, 0.93]	9860	0.13	0.63 [0.45, 0.81]	6190	0.30	0.59 [0.41, 0.77]	10510	0.0024	0.69 [0.57, 0.81]	6190	0.015	0.66 [0.53, 0.78]	9860
Creatinine mg/dl	0.00051	0.84 [0.72, 0.96]	0.95	0.00028	0.85 [0.74, 0.97]	1.05	0.0023	0.76 [0.63, 0.89]	0.95	0.026	0.69 [0.54, 0.84]	1.05	$5.2 \cdot 10^{-6}$	0.79 [0.70, 0.88]	0.95	0.000070	0.75 [0.65, 0.86]	1.05
Troponin ng/ml	0.0053	0.72 [0.56, 0.88]	0.005	0.0079	0.72 [0.55, 0.90]	0.005	0.0078	0.72 [0.57, 0.87]	0.005	0.020	0.69 [0.54, 0.85]	0.005	0.00010	0.73 [0.62, 0.83]	0.005	0.00027	0.72 [0.61, 0.83]	0.005
Ferritin ng/ml	0.064	0.72 [0.52, 0.91]	766	0.12	0.68 [0.47, 0.89]	530	0.0026	0.76 [0.62, 0.90]	1285	0.010	0.72 [0.58, 0.87]	1510	0.00023	0.75 [0.64, 0.86]	1285	0.0024	0.71 [0.59, 0.83]	1610

CRP = C-Reactive Protein; WBC= White blood cell count; LDH = Lactate Dehydrogenase; PCT = Procalcitonin; IL6 = Interleukin-6; AUC = area under the curve; CI = confidence interval

Table 3: Contingency table for high-risk and low-risk groups as defined by IL-6 and CRP in the validation cohort

Variable	Value	Mechanical ventilation		p-value
		No	Yes	
IL-6 at presentation	≤35	19	3	0.0030
	>35	11	16	
Maximal IL-6	≤80	25	5	0.0022
	>80	5	14	
CRP at presentation	≤32.5	19	1	0.00019
	>32.5	11	18	
Maximal CRP	≤97	23	3	0.00011
	>97	7	16	

Figure legends**Figure 1: Consort Diagram:**

Consort Diagram. DNR/DNI: do-not-resuscitate and do-not-intubate order.

Figure 2: IL-6 at presentation, maximal IL-6 levels before mechanical ventilation and ROC-analysis of different parameters in the evaluation and validation cohort

Box plots showing IL-6 levels at first assessment (A, B) and maximal IL-6 levels before mechanical ventilation (C, D) in the evaluation cohort and in the validation cohort; dashed lines represents the cutoff calculated from the evaluation cohort (IL-6 at initial assessment >35 pg/ml, maximal IL-6 >80 pg/ml). Mean \pm SD is shown. Receiver operating characteristic (ROC) curve of maximal follow-up levels before mechanical ventilation in the evaluation (E) and validation cohorts (F).

Figure 3: CRP levels at presentation and maximal CRP levels before mechanical ventilation

Box plot showing CRP levels at first assessment (A, B) and maximal IL-6 levels before mechanical ventilation (C, D) in the evaluation cohort and in the validation cohort; dashed lines represents the cutoff calculated from the training cohort (CRP at assessment >32.5 mg/l, maximal CRP>97 mg/l). Mean \pm SD is shown.

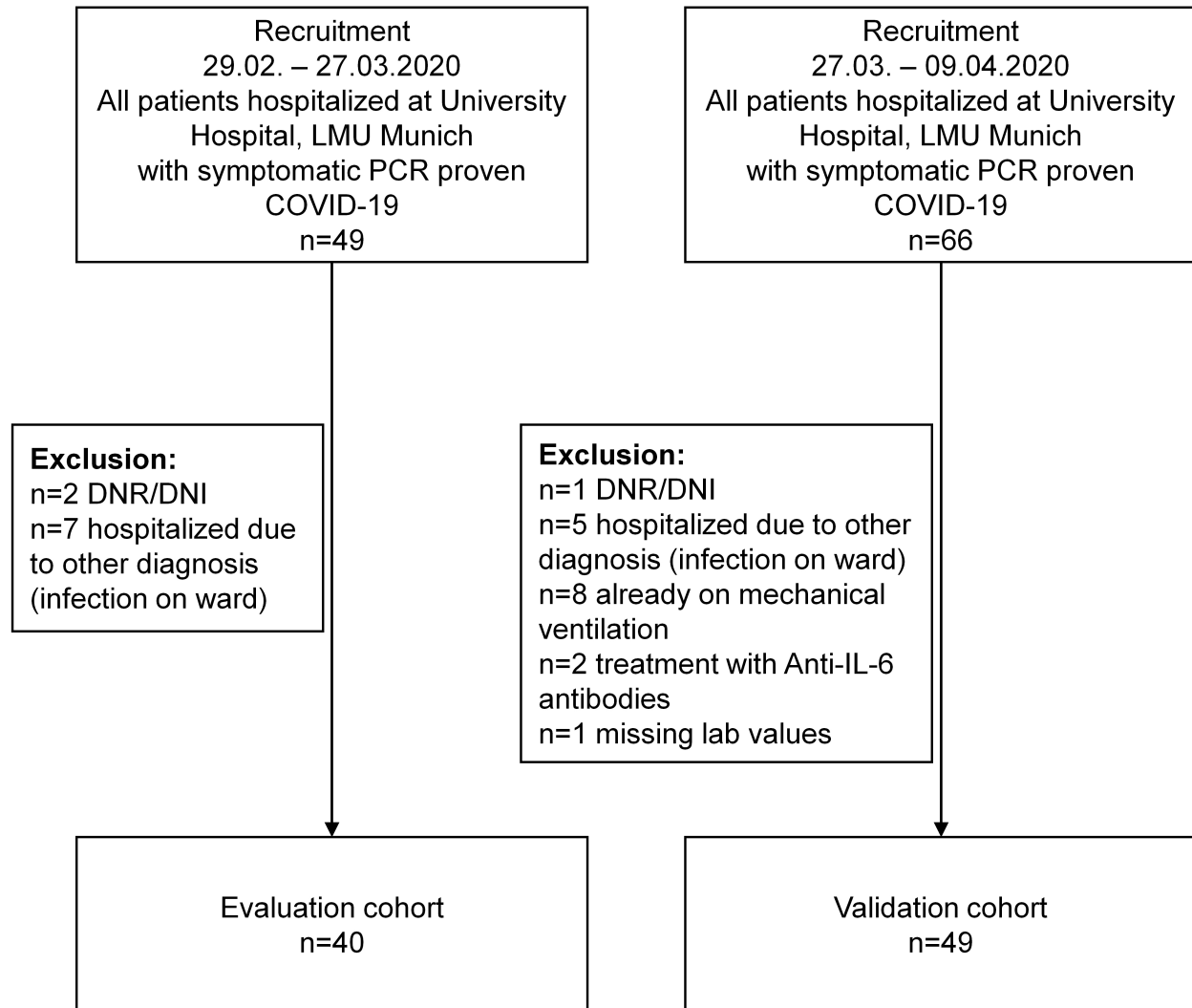
Figure 4: Cutoffs and predictive values of maximal IL-6 and CRP values in the combined cohort

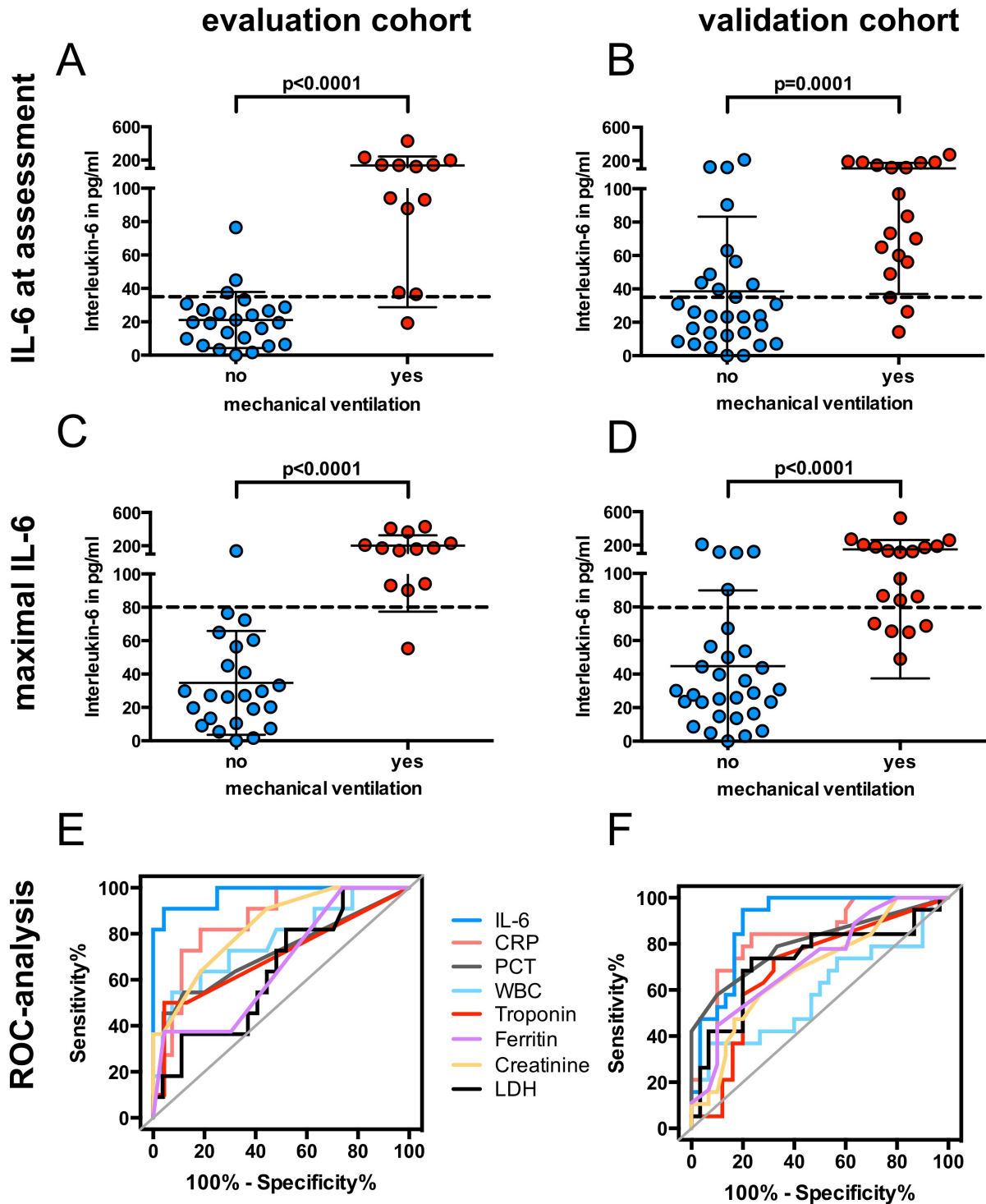
Box plots depicting the maximal values of IL-6 and CRP in the overall cohort (A, B); dashed line represents the validated cutoff; dotted line represents the calculated improved cutoff from all patients (applicable only for IL-6). Positive predictive value (PPV) and negative

predictive value (NPV) as a function of different cutoffs is shown for IL-6 (C) and CRP (D) values (dotted line represents cutoff for perfect NPV; dashed line represents cutoff for perfect PPV)

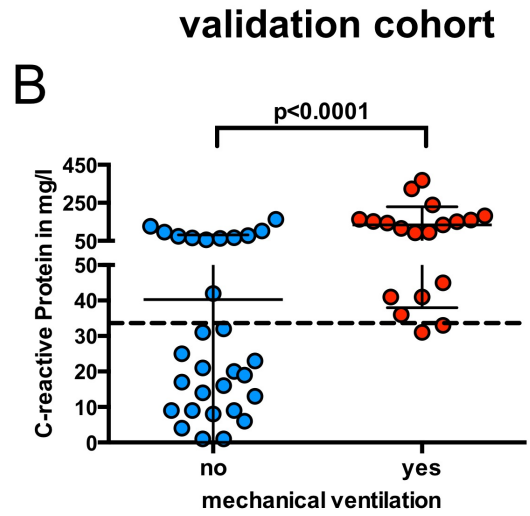
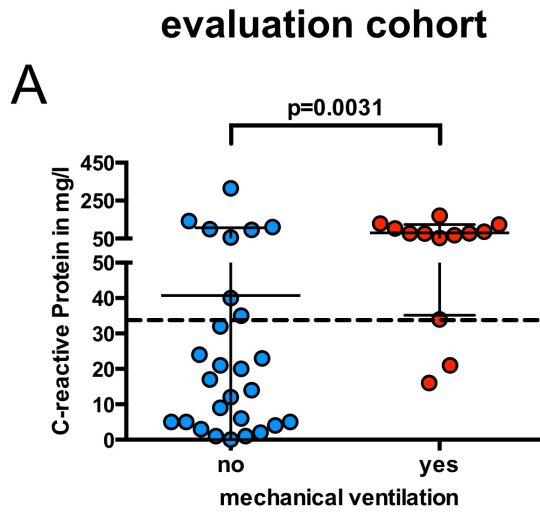
Figure 5: Time from exceeding the maximal cutoff value of IL-6 or CRP to intubation in the combined cohort

Box plot depicting the time from exceeding the IL-6 (>65 ng/ml) and CRP (>97 mg/l) cutoff to intubation in hours in the combined cohort. Median \pm min/max is shown.

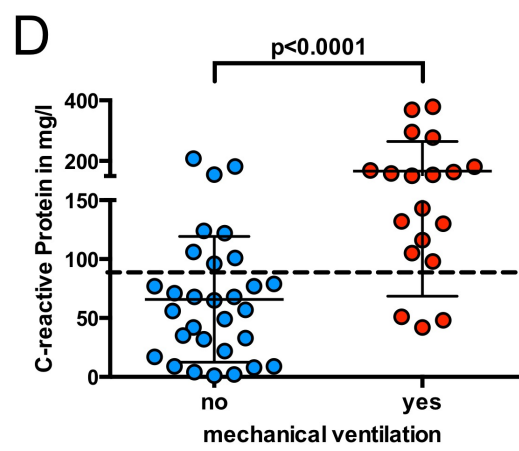
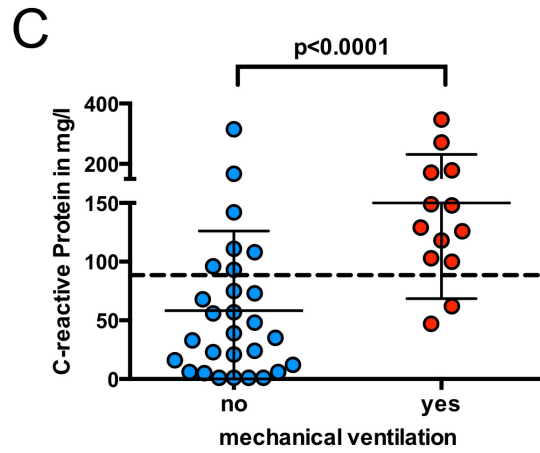


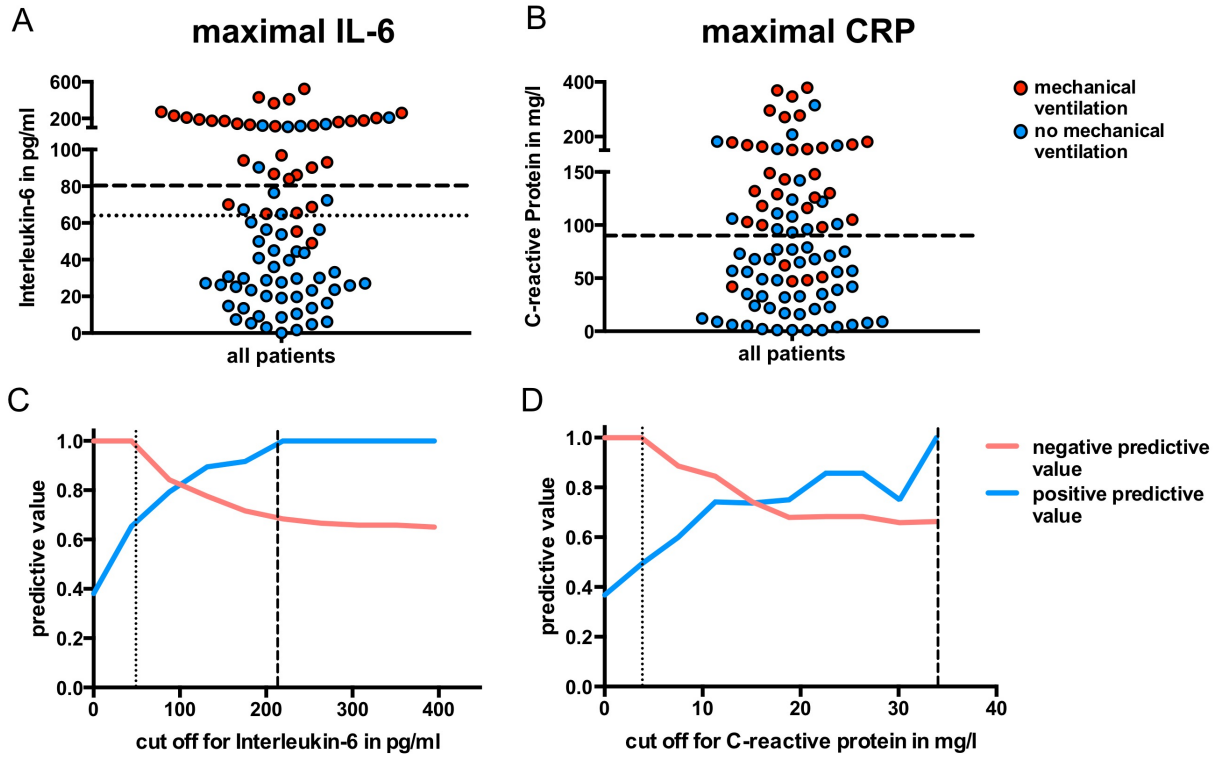


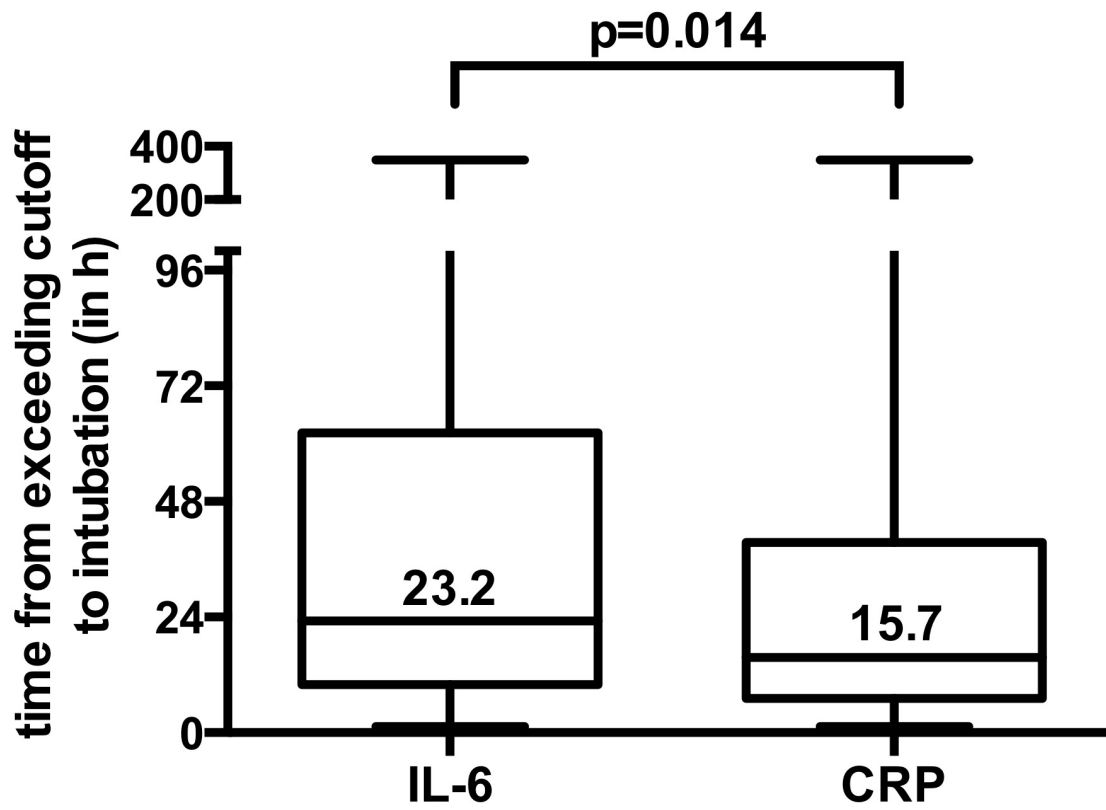
CRP at assessment



maximal CRP







Supplementary

**Elevated levels of interleukin-6 and CRP predicts the need for mechanical ventilation
in COVID-19**

Tobias Herold and, Vindi Jurinovic et. al.

Table E1: Evaluation Cohort

Variable	Evaluable	Median (range) / n (%)	Mechanical ventilation		p-value	q-value
			No (n = 27)	Yes (n = 13)		
Baseline Characteristics *						
Age (years)	40	57 (19 - 81)	54 (19 - 80)	64 (45 - 81)	0.15	0.29
Respiratory rate (/min)	34	18 (14 - 40)	18 (14 - 32)	23 (15 - 40)	0.066	0.14
Heart rate (/min)	32	81 (54 - 112)	77 (54 - 111)	94 (80 - 112)	0.0069	0.022
BMI	30	25.9 (19.0 – 45.7)	23.7 (19.0 – 34.7)	30.5 (24.8 – 45.7)	0.0030	0.014
Male gender	40	29 (72)	16 (59)	13 (100)	0.020	0.051
Any comorbidities	39	32 (82)	20 (77)	12 (92)	0.46	0.81
Hypertension	38	19 (50)	10 (40)	9 (69)	0.17	0.32
Diabetes mellitus	38	3 (8)	1 (4)	2 (15)	0.55	0.82
Coronary heart disease	36	3 (8)	3 (12)	0 (0)	0.52	0.82
Chronic obstructive lung disease	37	3 (8)	2 (8)	1 (8)	>0.99	>0.99
Computed Tomography [#]						
Consolidation	36	21 (58)	14 (61)	7 (54)	0.95	>0.99
Ground glass opacity	36	31 (86)	20 (87)	11 (85)	>0.99	>0.99
Bilateral infiltration	36	33 (92)	21 (91)	12 (92)	>0.99	>0.99
Scores [§]						
qSOFA score ¹	32	12 (37)	7 (32)	5 (50)	0.55	0.82
CURB-65 score ² ≥ 1	24	7 (29)	5 (31)	2 (25)	>0.99	>0.99
MuLBSTA score ³	29	9 (4 - 15)	9 (4 - 13)	7 (5 - 15)	0.89	>0.99
Laboratory parameters [#]	Evaluable	Median (range)	Mechanical ventilation		p-value	q-value
			No (n = 27)	Yes (n = 13)		
Lymphocyte count G/l	31	0.99 (0.45 – 2.50)	0.99 (0.45 – 1.80)	0.95 (0.57 – 2.50)	0.92	>0.99
CRP (mg/l)	40	28 (0 – 315)	17 (0 – 315)	77 (16 – 171)	0.0031	0.014
Bilirubin (mg/dl)	37	0.5 (0.2 – 1.9)	0.5 (0.2 – 1.2)	0.5 (0.4 – 1.9)	0.78	>0.99
WBC (G/l)	40	5.04 (2.12 - 308)	4.67 (2.12 – 10.8)	7.38 (4.67 - 308)	0.0028	0.014
LDH (U/l)	39	285 (153 - 1078)	258 (153 - 619)	381 (252 - 1078)	0.00062	0.0058
PCT (ng/ml)	38	0 (0 - 5)	0 (0 – 0.6)	0.1 (0 - 5)	0.0043	0.017
IL-6 (pg/ml)	37	27.1 (0 - 430)	19.6 (0 – 76.5)	121 (19.2 - 430)	0.000012	0.00034
Thrombocyte count (G/l)	40	161 (0.12 - 440)	162 (0.27 - 334)	160 (0.12 - 440)	0.74	>0.99
Troponin T (ng/ml)	34	0 (0 – 0.032)	0 (0 – 0.022)	0 (0 – 0.032)	0.0053	0.019
Creatinine (mg/dl)	40	0.9 (0.4 – 2.1)	0.9 (0.4 – 1.3)	1.0 (0.9 – 2.1)	0.00051	0.0058
D-Dimer	31	0.7 (0 – 2.9)	0.6 (0 – 2.2)	1.1 (0.6 – 2.9)	0.019	0.051
Ferritin (ng/ml)	31	626 (46 - 2153)	553 (46 - 1748)	810 (431 - 2153)	0.064	0.14

* respiratory rate and heart rate and BMI (Body mass index) were measured at admission; existing comorbidities were evaluated by patient history at admission; # CT-scans and laboratory parameters at admission; § scores were calculated at admission. CRP = C-Reactive Protein; WBC= White blood cell count; LDH = Lactate Dehydrogenase; PCT = Procalcitonin; IL6 = Interleukin-6; qSOFA score = predicts mortality in sepsis, CURB-65 score = predicts mortality in community-acquired pneumonia, MuLBSTA score = predicts mortality in patients with viral pneumonia; q-values represent the Benjamini-Hochberg adjusted p-values

Table E2: Validation cohort

Variable	Evaluable	Median (range) / n (%)	Mechanical ventilation		p-value	q-value
			No (n = 30)	Yes (n = 19)		
Baseline Characteristics *						
Age (years)	49	64 (18 - 84)	61 (18 - 84)	65 (46 - 81)	0.18	0.31
Respiratory rate (/min)	34	18 (11 - 40)	17 (13 - 39)	26 (11 – 40)	0.027	0.083
Heart rate (/min)	34	90 (64 - 130)	94 (74 - 130)	86 (64 - 107)	0.033	0.091
BMI	41	27.5 (18.1 – 36.2)	27.6 (18.1 – 36.2)	27.0 (18.4 – 34.7)	0.58	0.71
Male gender	49	33 (67)	17 (57)	16 (84)	0.091	0.21
Any comorbidities	48	38 (79)	23 (77)	15 (83)	0.85	0.96
Hypertension	48	26 (54)	15 (50)	11 (61)	0.65	0.76
Diabetes mellitus	48	10 (21)	6 (20)	4 (22)	>0.99	>0.99
Coronary heart disease	49	4 (8)	1 (3)	3 (16)	0.31	0.46
Chronic obstructive lung disease	49	6 (12)	5 (17)	1 (5)	0.46	0.61
Computed Tomography [#]						
Consolidation	42	25 (59)	16 (57)	9 (64)	>0.99	0.98
Ground glass opacity	42	41 (98)	27 (96)	14 (100)	>0.99	>0.99
Bilateral infiltration	42	37 (88)	23 (82)	14 (100)	0.24	0.37
Scores [§]						
qSOFA score ¹	39	18 (46)	6 (25)	12 (80)	0.0025	0.010
CURB-65 score ² ≥ 1	23	15 (65)	6 (55)	9 (75)	0.55	0.71
MuLBSTA score ³	39	11 (0 - 15)	10 (0 - 15)	13 (9 - 15)	0.038	0.096
Laboratory parameters [#]	Evaluable	Median (range)	Mechanical ventilation		p-value	q-value
			No (n = 30)	Yes (n = 19)		
Lymphocyte count G/l	36	0.80 (0.20 – 2.84)	0.73 (0.31 – 2.36)	0.94 (0.20 – 2.84)	0.43	0.60
CRP (mg/l)	49	42 (1 – 369)	22 (1 – 163)	134 (31 – 369)	0.000032	0.00068
Bilirubin (mg/dl)	47	0.5 (0.2 – 1.2)	0.4 (0.2 – 1.2)	0.6 (0.2 – 1.1)	0.16	0.30
WBC (G/l)	49	6.0 (0.15 – 25.8)	5.79 (1.92 – 12.4)	7.22 (0.15 – 25.8)	0.13	0.26
LDH (U/l)	49	336 (181 - 1121)	278 (181 - 502)	474 (240 - 1121)	0.00032	0.0022
PCT (ng/ml)	49	0 (0 – 2.3)	0 (0 – 0.3)	0.2 (0 – 2.3)	0.000073	0.00068
IL6 (pg/ml)	49	42.7 (0 - 272)	23.7 (0 - 209)	83.5 (14.2 - 272)	0.000072	0.00068
Thrombocyte count (G/l)	49	216 (93 - 450)	212 (112 - 383)	220 (93 - 450)	0.23	0.37
Troponin T (ng/ml)	44	0 (0 – 0.178)	0 (0 – 0.143)	0.022 (0 – 0.178)	0.0078	0.027
Creatinine (mg/dl)	49	0.9 (0.5 – 7.0)	0.9 (0.5 – 5.6)	1.1 (0.8 – 7.0)	0.0023	0.010
D-Dimer	45	0.8 (0 – 35.2)	0.6 (0 - 35)	0.9 (0 – 35.2)	0.11	0.24
Ferritin (ng/ml)	48	789 (30 - 3577)	508 (30 - 2578)	1692 (237 - 3577)	0.0026	0.010

* respiratory rate and heart rate and BMI (Body mass index) were measured at admission; existing comorbidities were evaluated by patient history at admission; # CT-scans and laboratory parameters at admission; § scores were calculated at admission. CRP = C-Reactive Protein; WBC= White blood cell count; LDH = Lactate Dehydrogenase; PCT = Procalcitonin; IL6 = Interleukin-6; qSOFA score = predicts mortality in sepsis, CURB-65 score = predicts mortality in community-acquired pneumonia, MuLBSTA score = predicts mortality in patients with viral pneumonia; q-values represent the Benjamini-Hochberg adjusted p-values

Supplementary Table E3: Comparison Evaluation and Validation cohort

Variable	Cohort		p-value
	Evaluation (n = 40)	Validation (n = 49)	
Baseline Characteristics *			
Age (years)	57 (19 - 81)	64 (18 - 84)	0.15
Respiratory rate (/min)	18 (14 - 40)	18 (11 - 40)	0.76
Heart rate (/min)	81 (54 - 112)	90 (64 - 130)	0.017
BMI	25.9 (19.0 – 45.7)	27.5 (18.1 – 36.2)	0.18
Male gender	29 (72)	33 (67)	0.77
Any comorbidities	32 (82)	38 (79)	0.95
Hypertension	19 (50)	26 (54)	0.87
Diabetes mellitus	3 (8)	10 (21)	0.17
Coronary heart disease	3 (8)	4 (8)	>0.99
Chronic obstructive lung disease	3 (8)	6 (12)	0.79
Computed Tomography [#]			
Consolidation	21 (58)	25 (60)	>0.99
Ground glass opacity	31 (86)	41 (98)	0.14
Bilateral infiltration	33 (92)	37 (88)	0.89
Scores [§]			
qSOFA score ¹	12 (37)	18 (46)	0.62
CURB-65 score ² ≥ 1	7 (29)	15 (65)	0.029
MuLBSTA score ³	9 (4 - 15)	11 (0 - 15)	0.13
Laboratory parameters [#]	Cohort		p-value
	Evaluation (n = 40)	Validation (n = 49)	
Lymphocyte count G/l	0.99 (0.45 – 2.5)	0.8 (0.2 – 2.84)	0.27
CRP (mg/l)	28 (0 – 315)	42 (1 – 369)	0.10
Bilirubin (mg/dl)	0.5 (0.2 – 1.9)	0.5 (0.2 – 1.2)	0.71
WBC (G/l)	5.04 (2.12 - 308)	6 (0.15 – 25.8)	0.47
LDH (U/l)	285 (153 - 1078)	336 (181 - 1121)	0.18
PCT (ng/ml)	0 (0 – 5)	0 (0 – 2.3)	0.32
IL-6 (pg/ml)	27.1 (0 - 430)	42.7 (0 - 272)	0.34
Thrombocyte count (G/l)	161 (0.12 - 440)	216 (93 - 450)	0.0084
Troponin T (ng/ml)	0 (0 – 0.032)	0 (0 – 0.178)	0.016
Creatinine (mg/dl)	0.9 (0.4 – 2.1)	0.9 (0.5 – 7.0)	0.82
D-Dimer	0.7 (0 – 2.9)	0.8 (0 – 35.2)	0.57
Ferritin (ng/ml)	626 (46 - 2153)	789 (30 - 3577)	0.20

* respiratory rate and heart rate and BMI (Body mass index) were measured at admission; existing comorbidities were evaluated by patient history at admission; [#] CT-scans and laboratory parameters at admission; [§] scores were calculated at admission. CRP = C-Reactive Protein; WBC= White blood cell count; LDH = Lactate Dehydrogenase; PCT =

Procalcitonin; IL6 = Interleukin-6; qSOFA score = predicts mortality in sepsis, CURB-65 score = predicts mortality in community-acquired pneumonia, MuLBSTA score = predicts mortality in patients with viral pneumonia; q-values represent the Benjamini-Hochberg adjusted p-values

Supplementary References

1. Seymour CW, Liu VX, Iwashyna TJ, Brunkhorst FM, Rea TD, Scherag A, *et al.* Assessment of Clinical Criteria for Sepsis: For the Third International Consensus Definitions for Sepsis and Septic Shock (Sepsis-3). *JAMA* **315**, 762-774 (2016).
2. Lim WS, Van der Eerden MM, Laing R, Boersma WG, Karalus N, Town GI, *et al.* Defining community acquired pneumonia severity on presentation to hospital: an international derivation and validation study. *Thorax* **58**, 377-382 (2003).
3. Guo L, Wei D, Wu Y, Zhou M, Zhang X, Li Q, *et al.* Clinical Features Predicting Mortality Risk in Patients With Viral Pneumonia: The MuLBSTA Score. *Front Microbiol* **10**, 2752 (2019).